The Enzymic Formation of Thiocyanate (SCN⁻) from a Precursor(s) in Brassica Species ROLF GMELIN and ARTTURI I. VIRTANEN

Laboratory of the Foundation for Chemical Research, Biochemical Institute, Helsinki, Finland

The chemical nature of the goitrogenic "Brassica factors" is still unclear to a great extent. This is partly due to the fact that several active factors may be involved, which are moreover formed only through enzymic reactions when plants are crushed. This has long been known in regard to the formation of the strongly goitrogenic thiooxazolidones occurring in the crushed and moistened seeds of many crucifers 1. Recently Virtanen et al.2, and Kreula and Kiesvaara 3 found the formation of vinyl-thiooxazolidone also in cabage, kale, rape, and other fodder plants belonging to the Cruciferae family.

The same authors 4 have also demonstrated the transfer of this substance to milk in very small amounts (about 0.05 % of the amount fed). These quantities are so small that they cannot be expected to have any goitrogenic effect in the milk. Kreula and Kiesvaara have given a detailed report on the methods used in these investigations. Somewhat later Altamura et al.5 independently also demonstrated the formation of vinyl-thiooxazolidone

in cabbage.

The goitrogenic effect of thiooxazolidones is due to the fact that they inhibit the synthesis of thyroid hormones. This effect cannot be overcome by high doses

of iodide.

Another type of goitrogenic substances belonging to the Brassica factors has a primary influence on the uptake of iodide by the thyroid gland. This influence can be prevented by increasing the amount of the iodide dose. Many pieces of information given in the literature 6,7 indicate that salts of thiocyanic acid (SCN) represent this type in crucifers. Jirousek's, who has investigated and reviewed the thiocyanate metabolism, found that in the animal organism SCN is formed endogenously from cyanides, nitriles, and sulphur-containing compounds, and that both the SCN brought exogenously into the organism and that formed endogenously in it have to be taken into consideration as goitrogenic factors. Michajlovskij and Langer ^{9,10} have performed systematic determinations on the SCN content in different vegetables. They found a particularly high SCN content in the press juice of different cabbage species (up to 50 mg %).
They used the term "präformiertes Rhodanid" for the SCN" present in food stuffs, obviously in contrast to the SCN formed endogenously.

Our investigations concerning the problem whether it is possible to make milk goitrogenic by feeding cows with plants, especially those belonging to the Cruciferae, have led to some new findings regarding the Brassica factors. These findings are briefly reported in the present paper.

An account was recently given about the formation of organic thiocyanates in some crucifers 11. Benzyl thiocyanate was found to be formed enzymatically from glucotropaeolin in Lepidium ruderale and allyl thiocyanate from sinigrin in Thlaspi arvense. It was found in this laboratory that after the injection of benzyl thiocyanate into rats, the SCN content has risen considerably in blood serum and in different organs. This finding led us to look for thiocyanate esters also in Brassica oleracea species. For the present we have, however, no indications for the formation of such esters in fresh cabbage species. On the other hand, the result of these investigations was the finding that free SCN is formed from glucosidic precursors, present in cabbage. The thiocyanate found by in cabbage. The thiocyanate found by Michajlovskij and Langer in the press juice of cabbage is thus not "preformed SCN"

Acta Chem. Scand. 14 (1960) No. 2

$$\begin{array}{c} \text{NOSO}_{3}^{-} & \xrightarrow{\text{myrosinase}} \\ \text{S-glucose} & \xrightarrow{\text{H}_{2}\text{O}} & [\text{R-N=C=S}] + \text{glucose} + \text{sulphate} \\ & \downarrow \\ \text{ROH}^{-} + \text{SCN}^{-} \end{array}$$

but is formed enzymatically from a glucosidic precursor(s) in cabbage. This fact was established when enzymes were destroyed in cabbage leaves by placing them intact in boiling methanol. The solution obtained was free from SCN. After addition of myrosinase solution to the purified extract SCN was formed, as determined by colour reactions for SCN and by paper chromatography.

The determination of SCN formed enzymatically in fresh plants was performed in the following way. 10 g of whole leaves of Brassica oleracea species were twice extracted by boiling for 15 min in 50 ml portions of methanol. The methanolic solutions were decanted. plant material was thoroughly ground in a mortar, and was once more extracted by boiling in 50 ml of 70 % methanol. The combined extracts were evaporated in vacuo. The residue was taken up in water, and the solution was treated with about 0.6 ml of 20 % lead acetate solution. Filtration and washing of the filter residue with water. Excess lead ions in the filtrate were precipitated by H2S. The filtrate was concentrated in vacuo to about 20 ml and brought to 25 ml (volumetric flask). 10 ml of this solution, 1 ml of myrosinase solution *, and 1 ml of phosphate buffer at pH 7 were incubated for 2 h at 37°. The solution was then brought to 100 ml. A control without addition of myrosinase solution was treated in the same way, and was used as a blank. 5 ml samples were mixed with 1 ml of Fe-reagent, measured at 500 m μ with a Klett-Summerson colorimeter, and the values compared with a standard SCN curve, essentially as in the method of Barker 13.

Table 1 shows the amounts of SCNformed in different cabbage species.

As is apparent from the list, varying amounts of SCN are formed in different cabbage species. The thiocyanate content of the press juices found by Langer and Michajlovskij corresponds to the amounts found by us.

Table 1. SCN content in cabbage species after myrosinase treatment.

CN formation per 00 g fresh plants
27-31 mg
10 mg
4 mg
4 mg
8.8 mg
2.5 mg
1.7 mg

- root

No formation of SCN was found in the crushed and moistened seeds of different cabbage species.

Attempts to isolate the glucosidic precursor from cabbage have been unsuccessful so far. The following findings can, however. be presented.

1. The precursor(s) of SCN decomposes with myrosinase so that SCN is liberated.

2. On spraying with AgNO₃ solution and heating, the precursor of SCN gives the same reaction on the paper chromatogram as mustard oil glucosides (gray-brown spot14)

3. On spraying with FeCl, solution, the precursor of SCN gives a brownish red spot after a short heating to 100°.

4. The position of the precursor on the paper chromatogram could be established furthermore either by spraying the paper first with myrosinase solution, and after standing for 1 h with FeCl₃ solution or by eluting the zones by the usual colour reactions for SCN after treatments of the eluates by myrosinase.

prepared according to Neuberg and Wagner 12. SCN originally present in the myrosinase solution, was removed by shaking the solution for 30 min with Dowex 2 imes 4 in chloride form.

5. In a butanol-acetic acid-water system, the R_F value ¹⁵ of the SCN precursor was 0.76, in a pyridine-amylalcohol-water sys-

tem it was 1.0.

Although the information about the chemical nature of the precursor is still scanty, we think it reasonable to publish a preliminary communication about our results at this stage. In our opinion the finding of the enzymatic formation of SCN is rather important, since it may help to shed light on many discrepancies about the Brassica factors.

If the SCN⁻ precursor is a mustard oil glucoside, as we suppose on the basis of the present indications, the formation of SCN⁻ according to the following reaction scheme

seems probable, (see p. 508).

This secondary decomposition of an isothiocyanate is no unknown reaction. Already in 1879 Will and Laubenheimer 16 have observed that p-hydroxybenzyl mustard oil (the isothiocyanate formed from the glucoside sinalbin, present in white mustard) is split almost quantitatively into

SCN by the action of alkali.

We have, however, found that considerable amounts of SCN are formed from glucosinalbin during the enzymatic cleavage even at pH 7. The initially present, pungent tasting p-hydroxybenzyl isothiocyanate decomposes gradually, loosing its irritating properties, into SCN and probably p-hydroxybenzyl alcohol. This secondary reaction seems to proceed at pH 7 at a slower rate than is the case with the SCN precursor of cabbage. Treatment of glucosinalbin with alkali, however, affords complete and fast fission, and permits a simple assay for the glucosinalbin content in the seeds of Sinapis alba by colorimetric SCN determination.

As long as the SCN precursor has not yet been isolated from Brassica oleracea species, the plants containing glucosinalbin (Sinapis species, Bunias species, Lepidium campestre etc.) may be used as suitable models for studies on the goitrogenic effect of the SCN precursor. Wagner-Jauregg and Koch 17 have for example observed a slight goitrogenic effect by application of myrosinase treated sinalbin solution to rabbits (sinalbin itself was without effect) but have not recognized or regarded SCN as the evidently responsible goitrogenic factor in this experiment.

Several thioglucosides were found in the limited number of cabbage species used so far for the attempted isolation of the SCN-precursor: glucoraphanin, sinigrin, and glu-

conapin in the fresh parts of Brassica oleracea var. gemmifera; glucoiberin and sinigrin in the fresh parts of marrow stem kale ("Chou Moellier"); glucoiberin and glucoraphanin in Brassica oleracea var. cretica. They were detected ¹⁴ directly by paper chromatography and comparison with authentic thioglucosides, or indirectly as thioureas derived from the enzymatically formed isothiocyanates by the method of Kjær and Rubinstein ¹⁸ in different solvent systems.

From 500 g fresh parts of Brassica oleracea var. gemmifera 15 mg of analytically pure sulphoraphan-phenylthiourea were

isolated (m. p. 145°C) *.

The presence of glucoiberin and glucoraphanin and their isothiocyanates, respectively, in cabbage species becomes of special interest since Bachelard and Trikojus ²⁰ have recently found that cheirolin (3-methylsulphonylpropyl isothiocyanate) has a significant thyreostatic effect in animals. Iberin (3-methylsulphinylpropyl isothiocyanate) and sulphoraphan (4-methylsulphinylbutyl isothiocyanate belong to the same isothiocyanate type as cheirolin, and may thus be considered as potential members in the group of "Brassica factors".

This investigation is a part of a research project under U.S. Public Law No. 480, 83rd Congress.

- Astwood, E. B., Greer, M. A. and Ettlinger, M. G. J. Biol. Chem. 181 (1949) 121.
- Virtanen, A. I., Kreula, M. and Kiesvaara, M. Acta Chem. Scand. 12 (1958) 580.
- Kreula, M. and Kiesvaara, M. Ibid. 13 (1959) 1375.
- Virtanen, A. I., Kreula, M. and Kiesvaara, M. *Ibid.* 13 (1959) 1043.
- Altamura, M. R., Long Jr., L. and Hasselström, T. J. Biol. Chem. 234 (1959) 1847.
- 6. Greer, M. A. Am. J. Clin. Nutrition 5 (1957)
- 7. Langer, P. Nature 185 (1960) 174.
- 8. Jirousek, L. Physiol. Bohemosloven. 5 (1956)
- Michajlovskij, N. and Langer, P. Hoppe-Seyler's Z. physiol. Chemie 312 (1958) 26; 317 (1959) 30.

^{*} The presence of glucoiberin and glucoraphanin in *Brassica oleracea* species was established two months ago by the isolation of iberin-phenylthiourea and sulforaphanphenylthiourea from a combined extract of *Brassica oleracea* var. *gemmijera* and var. *sabauda* by Procházka *et al.* ¹⁹

- Langer, P. and Michajlovskij, N. Ibid.
 312 (1958) 31.
 Gmelin, R. and Virtanen, A. I. Suomen Kemistiletti B 32 (1959) 236; Acta Chem. Scand. 13 (1959) 1474.
- 12. Neuberg, C. and Wagner, J. Biochem. Z. 174 (1926) 457. 13. Barker, H. J. Am. Med. Assoc. 106 (1936) 762.
- 14. Schultz, O. E. and Gmelin, R. Z. Natur-
- forsch. **7 b** (1952) 500. 15. Schultz, O. E. and Wagner, W. *Ibid.* **11 b** (1956) 73.
- 16. Will, H. and Laubenheimer, A. Ann. 199 (1879) 150.
- Wagner-Jauregg, Th. and Koch, J. Z. Naturforsch. 2 b (1947) 13.
 Kjær, A. and Rubinstein, K. Acta Chem. Scand. 7 (1953) 528.
- 19. Procházka, Z., Šanda, V. and Jirousek, L. Collection Czech. Chem. Commun. 24 (1959) 3606.
- Bachelard, H. S. and Trikojus, V. M. Nature 185 (1960) 80.

Received February 20, 1960.